Antitumor Agents. 125. New 4β-Benzoylamino Derivatives of 4'-O-Demethyl-4-desoxypodophyllotoxin and 4β-Benzoyl Derivatives of 4'-O-Demethylpodophyllotoxin as Potent Inhibitors of Human DNA Topoisomerase II

Xiao-Ming Zhou,² Zhe-Qing Wang,² Hong-Xing Chen,³ Yung-Chi Cheng,³ and Kuo-Hsiung Lee^{2,4}

Received February 2, 1992; accepted August 15, 1992

A series of 4β -benzoylamino (5–17) derivatives of 4'-O-demethyl-4-desoxypodophyllotoxin and 4β -benzoyl (18–20) derivatives of 4'-O-demethyl podophyllotoxin have been synthesized and evaluated for their inhibitory activity against the human DNA topoisomerase II as well as for their activity in causing cellular protein-linked DNA breakage. Compounds 5–13 and 15–17 are more potent than etoposide in causing DNA breakage, while compounds 9, 10, 13, 14, 16, and 20 are more active than etoposide in their inhibition of the human DNA topoisomerase II. The order for the enzyme inhibitory activity of the derivatives of 4'-O-demethyl-4-desoxypodophyllotoxin is 4β -arylamino $> 4\beta$ -benzylamino $> 4\beta$ -benzylamino.

KEY WORDS: etoposide; DNA topoisomerase II; amino analogues of etoposide; KB cells; cytotoxicity.

INTRODUCTION

Etoposide (VP-16; 1) (Fig. 1) is useful in the treatment of a variety of cancers, including small cell lung cancer, testicular cancer, lymphoma, and leukemia. In combination chemotherapy, it has been used successfully with cisplatin and several other antitumor drugs including cyclophosphamide, doxorubicin, and vincristine (2-4). Although it is widely used in the clinic, the use of 1 as a single agent has not proven curative in patients with germinal neoplasms. Prior treatment seems to be an important factor in determining the therapeutic outcome of VP-16-containing chemotherapy regimens (5,6). VP-16 is poorly soluble in water and has limited stability in physiological solution. It also becomes important to overcome the development of myelosuppression and resistance to 1 (4,6,7). All of these considerations have encouraged further synthesis of analogues of 1 with better pharmacological profiles.

It was reported that some amide derivatives of 4'-O-demethyl-4-desoxypodophyllotoxin (5–15; where R = 1

¹ For part 124, see Ref. 1.

P-NH-4,6-O-ethylideneglucopyranosyl) demonstrated a comparable antileukemic activity against L-1210 lymphocytic leukemia in vivo and less toxicity compared to 1 (8). Previously, we have demonstrated that some of the 4Bamino analogues, such as W-68 and NPF, are 40-fold more cytotoxic than 1 against P-388/adriamycin resistant cells (P-388/adr), which overexpressed MDR1 and decreased the content of topoisomerase II, whereas they exhibited almost the same activity against P-388 cells. In addition, NPF was found to be at least seven times less toxic in vivo compared to 1 (9,10). Since these 4β amino analogues possess potent activity in inhibiting DNA topoisomerase II and in causing cellular protein-linked DNA breakage (11-13), we have synthesized and evaluated a series of 4\beta-benzoylamino derivatives of 4'-O-demethyl-4-desoxypodophyllotoxin and 4βbenzoyl derivatives of 4'-O-demethylpodophyllotoxin as inhibitors of human DNA topoisomerase II and as part of our ongoing program aimed at elucidating the structure-activity relationships among these 4β-amino analogues of 1. The enzyme inhibitory activity of the amide derivatives of 4'-Odemethyl-4-desoxypodophyllotoxin mentioned above has not yet been reported. The DNA topoisomerase II has been proposed as a target enzyme of 1, as 1 inhibits the catalytic activity of DNA topoisomerase II by stabilizing a cleavable enzyme-DNA complex, in which DNA is cleaved and covalently linked to the enzyme (14–17).

MATERIALS AND METHODS

General Experimental Procedures

All melting points were taken on a Fischer-Johns melting-point apparatus and were uncorrected. IR spectra were recorded on a Perkin-Elmer 1320 spectrophotometer, and ¹H NMR spectra were obtained using a Bruker AC-300 NMR spectrometer. All chemical shifts are reported as parts per million from TMS. Elemental analyses were performed by Atlantic Microlab, Inc., Norcross, GA. Optical rotations were measured with a Rudolph Research autopol III polarimeter. Analytical thin-layer chromatography (TLC) was carried out on Merck precoated silica gel 60F-25 EM Kieselgel 60 (230- to 400-mesh ASTM). All new compounds were characterized by melting point, optical rotation, ¹H NMR, and IR spectra as well as elemental analyses.

Synthesis of Compounds 5-15

To a solution of an appropriate substituted benzoic acid (0.25 mmol) in THF (3 mL) was added DCC (57 mg, 0.28 mmol). After 10 min, compound 2 (100 mg, 0.25 mmol) was added to the mixture, and the mixture was stirred at room temperature overnight. The mixture was filtered and the filtrate was evaporated to give a crude product of 5–15. This product was purified by preparative TLC (chloroform:ethyl acetate:acetone:methanol = 100:5:5:5).

4'-O-Demethyl-4β-(benzoylamino)-4-desoxypodophyllotoxin (5). Yield, 73%; m.p., 213–214°C; crystals from chloroform-ethyl acetate; $[\alpha]_{D}^{25}$ -68° (c=0.25, acetone). IR (KBr) ν_{max} : 3500, 3300, 2910, 1750, 1720, 1610, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.78 (2H, d, J=7.4 Hz, H-2",6"), 7.57–7.45 (3H, m, H-3",4",5"), 6.83 (1H, s, H-5), 6.57 (1H, s, H-8), 6.33 (2H, s, H-2',6'), 6.27 (1H, d, J=6.8

Natural Products Laboratory, School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27514.

³ Department of Pharmacology, Yale University School of Medicine, New Haven, Connecticut 06520.

⁴ To whom correspondence should be addressed.

Antitumor Agents, 125 215

W - 68, R = NO₂

NPF, R = F

Fig. 1. Etoposide (VP-16; 1).

Hz, H-4), 6.01 and 5.99 (2H, s and s, OCH₂O), 5.45 (2H, brs, NH and OH-4'), 4.64 (1H, d, J = 4.3 Hz, H-1), 4.51 (1H, t, J = 9.2 Hz, H-11), 3.92 (1H, t, J = 9.2 Hz, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.06 (1H, m, H-3), and 2.93 (1H, dd, J = 14.2, 4.8 Hz, H-2). *Anal.* Calcd for C₂₈H₂₅NO₈ · ½H₂O: C, 65.63; H, 5.08; N, 2.73. Found: C, 65.65; H, 5.23; N, 2.71.

4'-O-Demethyl-4β-[(2"-hydroxybenzoyl)amino]-4-desoxypodophyllotoxin (6). Yield, 61%; m.p. 172–174°C; crystals from chloroform; [α]²⁵_D-69° (c=0.25, acetone). IR (KBr) $\nu_{\rm max}$: 3490, 3350, 3120, 2905, 1760, 1630, 1590, 1550, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.45 (1H, t, J=7.5 Hz, H-4"), 7.35 (1H, d, J=7.5 Hz, H-6"), 7.04 (1H, d, J=7.5 Hz, H-3"), 6.88 (1H, t, J=7.5 Hz, H-5"), 6.82 (1H, s, H-5), 6.58 (1H, s, H-8), 6.47 (1H, d, J=6.7 Hz, H-4), 6.40 (2H, s, H-2',6'), 6.01 and 6.00 (2H, s and s, OCH₂O), 5.44 (2H, brs, NH and OH-4'), 4.64 (1H, d, J=4.9 Hz, H-1), 4.49 (1H, t, H-11), 3.87 (1H, t, H-11), 3.76 (6H, s, OCH₃-3',5'), 3.05 (1H, m, H-3), and 2.96 (1H, dd, J=14.3, 4.9 Hz, H-2). Anal. Calcd for $C_{28}H_{25}NO_9 \cdot 1\frac{1}{2}H_2O$: C, 61.53; H, 5.13; N, 2.56. Found: C, 61.76; H, 5.28; N, 2.56.

4'-O-Demethyl-4β-[(2"-fluorobenzoyl)amino]-4-desoxypodophyllotoxin (7). Yield, 71%; m.p., 163–164°C; crystals from ethyl acetate-hexane; $[\alpha]^{25}_{\rm D}$ -58° (c=0.25, acetone). IR (KBr) $\nu_{\rm max}$: 3310, 3120, 2910, 1760, 1640, 1600, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.10 (1H, m, H-4"), 7.43 (1H, m, H-6"), 7.33 (1H, m, H-3"), 7.14 (1H, m, H-5"), 6.85 (2H, m, H-4,5), 6.57 (1H, s, H-8), 6.34 (2H, s, H-2',6'), 6.01 and 5.99 (2H, s and s, OCH₂O), 5.45 (2H, brs, NH and OH), 4.65 (1H, d, J=4.7 Hz, H-1), 4.50 (1H, t, H-11), 3.92 (1H, t, H-11), 3.81 (6H, s, OCH₃-3',5'), 3.00 (1H, m, H-3), and 2.95 (1H, dd, J=14.1, 4.9 Hz, H-2). Anal. Calcd for $C_{28}H_{24}NO_9$: C, 64.49; H, 4.61; N, 2.69. Found: C, 64.10; H, 4.99; N, 3.00.

4'-O-Demethyl-4β-[(3"-fluorobenzoyl)amino]-4-desoxypodophyllotoxin (8). Yield, 68%; m.p., 186–188°C; crystals from chloroform-ethyl acetate; $[\alpha]^{25}_D-62^\circ$ (c=0.25, acetone). IR (KBr) $\nu_{\rm max}$: 3490, 3300, 2900, 1750, 1630, 1570, 1500, and 1460 cm⁻¹. ¹H NMR (CDCl₃) δ 7.46 (3H, m, H-2",4",6"), 7.27 (1H, m, H-5"), 6.82 (1H, s, H-5), 6.57 (1H, s, H-8), 6.33 (3H, m, H-2',6' and H-4), 6.01 and 5.99 (2H, s and s, OCH₂O), 5.50 (2H, brs, NH and OH-4'), 4.63 (1H, d, J=5.7 Hz, H-1), 4.51 (1H, t, H-11), 3.89 (1H, m, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.05 (1H, m, H-3), and 2.93 (1H, dd, J=14.3, 4.8 Hz, H-2). Anal. Calcd for

 $C_{28}H_{24}NFO_8 \cdot \frac{1}{2}H_2O$: C, 63.39; H, 4.71; N, 2.64. Found: C, 63.01; H, 4.72; N, 2.92.

4'-O-Demethyl-4β-[(4"-fluorobenzoyl)amino]-4-desoxypodophyllotoxin (9). Yield, 69%; m.p., 242–244°C; crystals from chloroform–ethyl acetate; $[\alpha]^{25}_{D}$ -78° (c=0.25, acetone). IR (KBr) ν_{max} : 3410, 3120, 2910, 1760, 1630, 1590, 1510, and 1480 cm⁻¹; ¹H NMR (CDCl₃) δ 7.80 (2H, m, H-2",6"), 7.16 (2H, m, H-3",5"), 6.82 (1H, s, H-5), 6.57 (1H, s, H-8), 6.33 (2H, s, H-2',6'), 6.25 (1H, d, J=6.8 Hz, H-4), 6.01 and 6.00 (2H, s and s, OCH₂O), 5.43 (2H, brs, NH and OH-4'), 4.63 (1H, d, J=4.7 Hz, H-1), 4.50 (1H, t, H-11), 3.87 (1H, m, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.04 (1H, m, H-3), and 2.93 (1H, dd, J=14.2, 4.7 Hz, H-2). Anal. Calcd for $C_{28}H_{24}NFO_8$: C, 64.49; H, 4.61; N, 2.69. Found: C, 64.31; H, 5.09.

4'-O-Demethyl-4β-[(4"-acetoxybenzoyl)amino]-4-desoxypodophyllotoxin (10). Yield, 51%; m.p., 175–176°C; crystals from ethyl acetate–hexane; $[\alpha]^{25}_{\rm D}$ -61° (c=0.25, acetone). IR (KBr) $\nu_{\rm max}$: 3350, 3100, 2980, 1760, 1740, 1620, 1590, 1505, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.12 (2H, d, J=8.7 Hz, H-2",6"), 7.19 (2H, d, J=8.7 Hz, H-3",5"), 6.82 (1H, s, H-5), 6.57 (1H, s, H-8), 6.33 (2H, s, H-2',6'), 6.25 (1H, d, J=6.8 Hz, H-4), 6.01 and 6.00 (2H, s and s, OCH₂O), 5.43 (2H, s, NH and OH-4'), 4.63 (1H, d, J=4.7 Hz, H-1), 4.50 (1H, q, H-11), 3.86 (1H, m, H-11), 3.76 (6H, s, OCH₃-3',5'), 3.04 (1H, m, H-3), 2.96 (1H, dd, J=14.3, 4.7 Hz, H-2), and 2.33 (3H, s, CH₃CO). Anal. Calcd for $C_{30}H_{27}NO_{10}$: C, 64.17; H, 4.81; N, 2.50. Found: C, 64.01; H, 4.99; N, 2.44.

4'-O-Demethyl-4β-[(4"-acetylbenzoyl)amino]-4-desoxypodophyllotoxin (11). Yield, 70%; m.p., 178–180°C (dec); crystals from ethyl acetate–hexane; $[\alpha]^{25}_{D}$ -68° (c=0.25, acetone). IR (KBr) ν_{max} : 3500, 3350, 2920, 1770, 1680, 1640, 1600, 1520, and 1480 cm⁻¹. ¹H NMR (CDCl₃) δ 8.05 (2H, d, J=8.2 Hz, H-3",5"), 7.87 (2H, d, J=8.2 Hz, H-2",6"), 6.83 (1H, s, H-5), 6.58 (1H, s, H-8), 6.36 (3H, m, H-2',6' and H-4), 6.02 and 6.00 (2H, s and s, OCH₂O), 5.45 (2H, s, NH and OH-4'), 4.64 (1H, d, J=4.8 Hz, H-1), 4.51 (1H, t, H-11), 3.89 (1H, m, H-11), 3.73 (6H, s, OCH₃-3',5'), 3.06 (1H, m, H-3), 2.94 (1H, dd, J=14.2, 4.8 Hz, H-2), and 2.66 (3H, s, CH₃CO). Anal. Calcd for $C_{30}H_{27}NO_9 \cdot \frac{1}{2}H_2O$: C, 64.98; H, 5.05; N, 2.53. Found: C, 64.95; H, 5.09; N, 2.75.

4'-O-Demethyl-4β-[(3"-cyanobenzoyl)amino]-4-desoxypodophyllotoxin (12). Yield, 68%; m.p., 190–192°C; crystals from ethyl acetate–hexane; $[\alpha]^{25}_{D}$ -62° (c=0.25, acetone). IR (KBr) ν_{max} : 3300, 3100, 2910, 2200, 1760, 1640, 1590, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.06 (2H, m, H-2",6"), 7.84 (1H, d, J=7.5 Hz, H-4"), 7.62 (1H, t, J=7.5 Hz, H-5"), 6.82 (1H, s, H-5), 6.58 (1H, s, H-8), 6.38 (1H, d, J=6.7 Hz, H-4), 6.33 (2H, s, H-2',6'), 6.02 and 6.00 (2H, s and s, OCH₂O), 5.45 (2H, brs, NH and OH-4'), 4.64 (1H, d, J=4.8 Hz, H-1), 4.50 (1H, t, H-11), 3.87 (1H, t, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.07 (1H, m, H-3), and 2.94 (1H, dd, J=14.3, 4.9 Hz, H-2). Anal. Calcd for $C_{29}H_{24}N_2O_8 \cdot \frac{1}{2}H_2O$: C, 64.81; H, 4.66; N, 5.21. Found: C, 64.89; H, 4.72; N, 5.21.

4'-O-Demethyl-4β-[(4"-cyanobenzoyl)amino]-4-desoxypodophyllotoxin (13). Yield, 73%; m.p., 198–202°C; crystals from ethyl acetate–chloroform; [α]²⁵_D-64° (c=0.25, acetone). IR (KBr) $\nu_{\rm max}$: 3320, 3100, 2980, 2200, 1760, 1640, 1600, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.89 (2H, d, J=8.5 Hz, H-3",5"), 7.77 (2H, d, J=8.5 Hz, H-2",6"), 6.81

(1H, s, H-5), 6.58 (1H, s, H-8), 6.33 (3H, m, H-2',6' and H-4), 6.02 and 6.00 (2H, s and s, OCH₂O), 5.44 (2H, s, NH and OH-4'), 4.64 (1H, d, J = 5.0 Hz, H-1), 4.50 (1H, q, H-11), 3.83 (1H, q, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.06 (1H, m, H-3), and 2.91 (1H, dd, J = 14.3, 5.0 Hz, H-2). *Anal.* Calcd for $C_{29}H_{24}N_2O_8 \cdot \frac{1}{2}H_2O$: C, 64.81; H, 4.66; N, 5.21. Found: C, 64.77; H, 4.71; N, 5.21.

4'-O-Demethyl-4β-[(3"-nitrobenzoyl)amino]-4-desoxypodophyllotoxin (14). Yield, 80%; m.p., 194–195°C; crystals from ethyl acetate–chloroform; $[\alpha]^{25}_{D}$ -46° (c=0.25, acetone). IR (KBr) ν_{max} : 3350, 3120, 1750, 1640, 1600, 1510, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.58 (1H, s, H-2"), 8.38 (1H, d, J=7.4 Hz, H-4"), 8.21 (1H, d, J=7.5 Hz, H-6"), 7.26 (1H, t, H-5"), 6.82 (1H, s, H-5), 6.54 (2H, m, H-4,8), 6.32 (2H, s, H-2',6'), 6.00 (2H, s, OCH₂O), 5.45 (2H, brs, NH and OH-4'), 4.62 (1H, d, J=4.7 Hz, H-1), 4.49 (1H, t, H-11), 3.87 (1H, t, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.08 (1H, m, H-3), and 2.96 (1H, dd, J=14.4, 4.7 Hz, H-2). Anal. Calcd for $C_{28}H_{24}N_2O_{10} \cdot \frac{1}{2}H_2O$: C, 60.32; H, 4.76; N, 5.03. Found: C, 60.71; H, 4.51; N, 4.96.

4'-O-Demethyl-4β-[(4"-nitrobenzoyl)amino]-4-desoxypodophyllotoxin (15). Yield, 69%; m.p., 204–206°C; crystals from ethyl acetate–chloroform; $[\alpha]^{25}_{D}$ -52° (c=0.25, acetone). IR (KBr) ν_{max} : 3450, 3150, 2980, 1760, 1640, 1600, 1540, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.31 (2H, d, J=8.7 Hz, H-3",5"), 7.97 (2H, d, J=8.7 Hz, H-2",6"), 6.83 (1H, s, H-5), 6.58 (1H, s, H-8), 6.56 (1H, d, J=6.5 Hz, H-4), 6.32 (2H, s, H-2',6'), 6.02 and 5.99 (1H, s and s, OCH₂O), 5.45 (2H, brs, NH and OH-4'), 4.63 (1H, d, J=4.8 Hz, H-1), 4.51 (1H, t, H-11), 3.88 (1H, t, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.05 (1H, m, H-3), and 2.95 (1H, dd, J=14.3, 5.0 Hz, H-2). Anal. Calcd for $C_{28}H_{24}N_2O_{10} \cdot \frac{1}{2}H_2O$: C, 60.32; H, 4.76; N, 5.03. Found: C, 60.33; H, 4.78; N, 4.76.

Synthesis of Compounds 16 and 17

To a solution of 14 or 15 (25 mg, 0.05 mmol) in ethyl acetate (3.0 mL) was added 10% palladium on activated carbon (3.0 mg). The reaction mixture was stirred under hydrogen for 2 hr at room temperature under atmospheric pressure. The catalyst was filtered off and the filtrate was evaporated to give a crude product. The crude product was purified by preparative TLC (chloroform:acetone:ethyl acetate:methanol = 100:5:5:3) to yield 16 or 17, respectively.

4'-O-Demethyl-4 β -[(3'-aminobenzoyl)amino]-4-desoxypodophyllotoxin (16). Yield, 95%; m.p., 180-182°C; crystals from ethyl acetate-chloroform; [α]²⁵_D-53° (c = 0.25, acetone). IR (KBr) ν_{max} : 3360, 3120, 2920, 1760, 1640, 1600, 1570, and 1460 cm⁻¹. ¹H NMR (CDCl₃) δ 7.20 (1H, t, J = 7.7 Hz, H-5"), 7.11 (1H, s, H-2"), 7.02 (1H, d, J = 7.7 Hz, H-6"), 6.82 (2H, m, H-4" and H-5), 6.55 (1H, s, H-8), 6.32 (2H, s, H-2',6'), 5.98 (1H, d, J = 5.1 Hz, H-4), 5.99 and 5.97 (2H, s and s, OCH₂O), 5.40 (2H, brs, NH and OH-4'), 4.61 (1H, d, J = 4.6 Hz, H-1), 4.48 (1H, t, H-11), 3.85 (1H, m, H-11), 3.79 (6H, s, OCH₃-3',5'), 3.01 (1H, m, H-3), and 2.90 (1H, dd, J = 14.3, 4.6 Hz, H-2). Anal. Calcd for $C_{28}H_{26}N_2O_8$: C, 64.86; H, 5.02; N, 5.41. Found: C, 64.73; H, 5.24; N, 5.25.

4'-O-Demethyl-4β-[(4"-aminobenzoyl)amino]-4desoxypodophyllotoxin (17). Yield, 69%; m.p., 204–206°C; crystals from ethyl acetate–chloroform; $[\alpha]^{25}_{D}$ -56° (c = 0.25, acetone). IR (KBr) ν_{max} : 3450, 3350, 3150, 2960, 1760, 1630, 1600, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.60 (2H, d, J=8.5 Hz, H-2",6"), 6.82 (1H, s, H-5), 6.67 (2H, d, J=8.5 Hz, H-3",5"), 6.33 (2H, s, H-2',6'), 6.10 (1H, d, J=6.8 Hz, H-4), 6.00 and 5.98 (2H, s and s, OCH₂O), 5.43 (2H, brs, NH and OH-4'), 4.63 (1H, d, J=4.6 Hz, H-1), 4.49 (1H, t, H-11), 3.91 (1H, t, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.03 (1H, m, H-3), and 2.93 (1H, dd, J=14.2, 4.6 Hz, H-2). *Anal.* Calcd for C₂₈H₂₆N₂O₈ · $\frac{1}{2}$ H₂O: C, 63.76; H, 5.12; N, 5.31. Found: C, 64.02; H, 5.30; N, 5.12.

Synthesis of Compounds 18-20

To a solution of appropriate benzoic acids (0.34 mmol) in pyridine was added phenyl sulfonyl chloride (72 μL, 0.56 mmol) at 0°C. After 10 min the cooling bath was removed and the temperature was raised to 20–25°C. 4′-O-Demethyl-4-O-carbobenzoxyepipodophyllotoxin (3) (150 mg, 0.28 mmol) was added to the reaction mixture. After the mixture was stirred overnight, it was poured into ice water, extracted with dichloromethane, dried (Na₂SO₄), and evaporated. The resulting product was purified by preparative TLC (chloroform:acetone:ethyl acetate = 100:5:5) to give 4′-O-demethyl-4′-O-CBZ protected 18–20, respectively. These compounds were then subjected to catalytic hydrogenation in 4 mL of ethyl acetate with 30 mg of palladium on activated carbon for 2 hr at room temperature to yield 18–20, respectively.

4'-O-Demethyl-4β-benzoylpodophyllotoxin (18). Yield, 90%; m.p., 213–214°C; crystals from ethyl acetate-chloroform; $[\alpha]^{25}_{D}$ -79° (c=0.25, acetone). IR (KBr) ν_{max} : 3120, 1760, 1690, 1600, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 8.03 (2H, d, J=7.2 Hz, H-2",6"), 7.60 (1H, t, J=7.2 Hz, H-4"), 7.47 (2H, t, J=7.2 Hz, H-3",5"), 6.96 (1H, s, H-5), 6.60 (1H, s, H-8), 6.41 (1H, d, J=3.5 Hz, H-4), 6.27 (2H, s, H-2',6'), 6.01 and 5.97 (2H, s and s, OCH₂O), 5.43 (1H, s, OH-4'), 4.73 (1H, d, J=4.9 Hz, H-1), 4.40 (1H, t, H-11), 3.98 (1H, t, H-11), 3.81 (6H, s, OCH₃-3',5'), 3.36 (1H, dd, J=14.1, 4.9 Hz, H-2), and 3.10 (1H, m, H-3). Anal. Calcd for C₂₈H₂₄O₉: C, 66.67; H, 4.76. Found: C, 66.67; H, 4.76

4'-O-Demethyl-4β-(3"-aminobenzoyl)podophyllotoxin (19). Yield, 89%; m.p., 228–229°C; crystals from ethyl acetate-chloroform; $[\alpha]^{25}_{D}$ -78° (c=0.25, acetone). IR (KBr) ν_{max} : 3460, 3360, 2950, 1760, 1695, 1600, 1495, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.40 (1H, d, J=8.0 Hz, H-6"), 7.31 (1H, s, H-2"), 7.23 (1H, t, J=8.0 Hz, H-5"), 6.95 (1H, s, H-5), 6.90 (1H, d, J=8.0 Hz, H-4"), 6.59 (1H, s, H-8), 6.37 (1H, d, J=3.6 Hz, H-4), 6.34 (2H, s, H-2',6'), 6.01 and 5.97 (2H, s and s, OCH₂O), 5.45 (1H, s, OH-4'), 4.72 (1H, d, J=5.0 Hz, H-1), 4.39 (1H, t, H-11), 3.97 (1H, t, H-11), 3.81 (6H, s, OCH₃-3',5'), 3.34 (1H, dd, J=14.1, 5.0 Hz, H-2), and 3.08 (1H, m, H-3). Anal. Calcd for $C_{28}H_{25}NO_9$: C, 64.74; H, 4.82; N, 2.70. Found: C, 64.58; H, 4.95; N, 2.65.

4'-O-Demethyl-4β-(4"-aminobenzoyl)podophyllotoxin (20). Yield, 84%; m.p., 228–229°C; crystals from ethyl acetate–chloroform; $[\alpha]^{25}_{D}$ -62° (c=0.25, acetone). IR (KBr) ν_{max} : 3460, 3120, 2940, 1760, 1660, 1590, 1500, and 1470 cm⁻¹. ¹H NMR (CDCl₃) δ 7.83 (2H, d, J=8.5 Hz, H-2",6"), 6.95 (1H, s, H-5), 6.64 (2H, d, J=8.5 Hz, H-3",5"), 6.58 (1H, s, H-8), 6.34 (3H, brs, H-2',6' and H-4), 6.00 and 5.97

Antitumor Agents, 125 217

(2H, s and s, OCH₂O), 5.44 (1H, s, OH-4'), 4.71 (1H, d, J = 4.9 Hz, H-1), 4.38 (1H, t, H-11), 3.97 (1H, t, H-11), 3.80 (6H, s, OCH₃-3',5'), 3.34 (1H, dd, J = 14.1, 4.9 Hz, H-2), and 3.06 (1H, m, H-3). *Anal*. Calcd for $C_{28}H_{25}NO_9 \cdot \frac{1}{3}H_2O$: C, 64.00; H, 4.89; N, 2.67. Found: C, 64.16; H, 5.01; N, 2.64.

RESULTS AND DISCUSSION

As shown in Scheme I, 4β -(substituted benzoylamino)-4'-O-demethyl-4-desoxypodophyllotoxins (5–15) were synthesized by amidation of the appropriate substituted benzoic acids with 4'-O-demethyl- 4β -amino-4-desoxypodophyllotoxin (2), which was obtained from podophyllotoxin as described previously (13) (Table I). The yields in this synthesis

are in the range of 61–73% calculated from 2. In the presence of DCC, the acid selectively reacts with the amino group to form the amide rather than with a hydroxyl group to form the ester (18). Compounds 16 and 17 were prepared from their corresponding 14 and 15 by catalytic hydrogenation with PdC. Treatment of 3, prepared previously from 4'-O-demethylepipodophyllotoxin (19), with a substituted benzoic acid in the presence of benzenesulfonyl chloride (20) afforded the ester (4). Removal of the phenolic protecting group of 4 by catalytic hydrogenation furnished 18, 19, and 20. The yields in these syntheses are in the range of 84–90%, calculated from 3.

The selection of F, OH, OCOCH₃, COCH₃,CN,NO₂, and NH₂ as substituents of the amide or ester phenyl ring is

Table I. Biological Evaluation of 4β-(Substituted Benzoyl)-Amino-4'-O-demethyl-4-desoxypodophyllotoxins and 4β-(Substituted Benzoyl)-4'-O-Demethylpodophyllotoxins

Compound	R	Cyto- toxicity, ID ₅₀ KB (μΜ) ^a	Inhibition of DNA toposomerase II activity, ID ₅₀ (µM) ^b	Cellular protein– DNA complex formation (%) 10 µM
1	H ₃ C 0 0 0 0 0 0	0.20	50	100
5	O 1 HNC - 2" 4"	1.27	>50	177
6	HNC OH	<1.92	50	160
7	HNC F	2.50	100	128
8	HNC F	1.92	>50	116
9	O HNC F	0.65	25	117
10	HNC — OCCH ₃	1.09	25	137
11	HNC -CCH ₃	1.83	50	124
12	HNC CN	1.89	50	149
13	HNC CN	0.19	25	159
14	HNC NO ₂	0.60	10	86

15	HNC NO ₂	0.73	50	160
16	HNC — NH ₂	0.81	25	149
17	HNC -NH ₂	1.16	50	120
18		< 0.20	50	94
19	oc-NH ₂	1.56	50	100
20	$OC \longrightarrow NH_2$	1.19	25	94

^a ID₅₀ is the concentration of drug that affords 50% reduction in cell number after a 3-day incubation.

based on the fact that these phenyl substituents contribute to the enhanced inhibitory activity against DNA topoisomerase II and ability to cause protein-linked DNA breakage as observed in our previous studies on 4β-amino analogues (9–13). As illustrated in Table I, all 4β-substituted benzoyl amides and benzoyl esters (5-20) showed comparable or superior activity to 1 in inhibiting the human DNA topoisomerase II and in causing cellular protein-DNA strand breakage, except for 7, in which its inhibitory activity against the enzyme is less than that of 1. The most active compounds are 12, 13, 15, and 16. However, the enzyme inhibitory activity for these amides is less than that for their corresponding 4βarylamino (12) and 4β-benzylamino compounds (13) which possess the same substituents. A comparison of the amides with the esters showed that the amides are more potent than their corresponding esters. This result is in agreement with our previous observation that the 4β-nitrogen substituted aryl compounds are, in general, more potent than their corresponding 4β-etheral derivatives in inhibiting the human DNA topoisomerase II and in causing cellular protein-linked DNA breakage (11–13). There is a lack of correlation among the ability of compounds in causing protein linked DNA breaks and in producing cytotoxicity (KB).

ACKNOWLEDGMENTS

The authors thank Mike Fisher of the Cancer Research Center, UNC—Chapel Hill, for KB cell culture assay. This work was supported by American Cancer Society Grants CH-370 and DHP 13E (K.-H. Lee) and National Cancer Institute Grant CA 44358 (Y.-C. Cheng).

REFERENCES

1. Z. Q. Wang, H. Hu, H. X. Chen, Y. C. Cheng, and K. H. Lee.

b Each compound was examined with five concentrations, 10, 20, 25, 50, and 100 μM. The ID₅₀ value was established on the basis of the degree of inhibition at these five concentrations.

- Antitumor agents 124. New 4β-substituted aniline derivatives of 6,7-O,O-demethylene-4'-O-demethyl-podophyllotoxin and related compounds as potent inhibitors of human DNA topoisomerase II. *J. Med. Chem.* 35:871–877 (1992).
- P. O'Dwyer, B. Leyland-Jones, M. T. Alonso, S. Marsoni, and R. E. Wittes. Drug therapy etoposide (VP-16-213) current status of an active anticancer drug. N. Engl. J. Med. 312:692-700 (1985).
- 3. B. F. Issell, F. M. Muggia, and S. K. Carter. *Etoposide [VP-16]: Current Status and New Developments*, Academic Press, Orlando, FL, 1984, pp. 1-353.
- J. M. S. van Maanen, J. Retel, J. de Vries, and H. M. Pinedo. Mechanism of action of antitumor drug etoposide: A review. J Natl. Cancer Inst. 80(19):1526-1533 (1988).
- F. Cavalli, A. Goldhirsch, and R. Joss. In B. F. Issel, F. M. Muggia, and S. K. Carter (eds.), Etoposide (VP-16): Current Status and New Developments, Academic Press, New York, 1985, pp. 163-169.
- J. D. Hainsworth, S. D. Williams, L. H. Einhorn, R. Birch, and F. A. Greco. Successful treatment of resistant germinal neoplasms with VP-16 and cisplatin: Results of a Southeastern Cancer Study Group trial. J. Clin. Oncol. 3:666-671 (1985).
- J. C. Shah, J. R. Chen, and D. Chow. Preformulation study of etoposide: Identification of physiocochemical characteristics responsible for the low and erratic oral bioavailability of etoposide. *Pharm. Res.* 6:408-412 (1989).
- 8. K. Kurabayashi, H. Saito, and H. Machida. N-Glycosylpodophyllotoxins as anticancer agents and *P*-aminobenzoic acid N-4,6-O-ethylidene-D-glucopyranoside as their intermeniates. *Jpn. Kokai Tokkyo Koho. Jp* 01, 197, 486 [89, 197, 486] (Cl. C07D493/04), 9 Aug. 1989, Appl. 88/21, 915, 3 Feb. 1988.
- S. Y. Liu, B. D. Hwang, M. Haruma, Y. Imakura, K. H. Lee, and Y. C. Cheng. Podophyllotoxin agents: Effects on DNA topoisomerase II, tubulin polymerization, human tumor KB Cells, and their VP-16-resistant variants. *Mol. Pharmacol.* 36: 78-82 (1989).
- H. X. Chen, J. Y. Chang, Z. Q. Wang, K. H. Lee, and Y. C. Cheng. *In vitro* and *in vivo* studies on the toxicity and antitumor activity of etoposide (VP-16) and its analogs. Am. Assoc. Cancer Res. Annu. Meet., Washington, D.C., May 14-18, 1991, abstr., 2144.

- K. H. Lee, S. A. Beers, M. Mori, Z. Q. Wang, Y. H. Kuo, L. Li, S. Y. Liu, J. Y. Chang, F. S. Han, and Y. C. Cheng. Antitumor agents. 111. New 4-hydroxylated and 4-halogenated anilino derivatives of 4'-demethylepipodophyllotoxin as potent inhibitors of human DNA topoisomerase II. J. Med. Chem. 33:1364-1368 (1990).
- Z. Q. Wang, Y. H. Kuo, D. Schnur, J. P. Bowen, S. Y. Liu, F. S. Han, J. Y. Chang, Y. C. Cheng, and K. H. Lee. Antitumor agents. 113. New 4β-arylamino derivatives of 4'-O-demethylepipodophyllotoxin and related compounds as potent inhibitors of human DNA topoisomerase II. J. Med. Chem. 33:2660-2666 (1990).
- X. M. Zhou, Z. Q. Wang, J. Y. Chang, Y. C. Chen, and K. H. Lee. Antitumor agents. 120. New 4β-substituted benzylamine and benzyl ether derivatives of 4'-O-demethylepipodophyllotoxin as potent inhibitors of human DNA topoisomerase II. J. Med. Chem. 34:3346–3350 (1991).
- 14. R. B. Lock and W. E. Ross. DNA topoisomerases in cancer therapy. *Anti-Cancer Drug Design* 2:151-164 (1987).
- G. L. Chen, L. Yang, T. C. Rowe, B. D. Halligan, K. Tewey, and L. Liu. Nonintercalative antitumor drugs interfere with the breakage-reunion reaction of mammalian DNA topoisomerase II. J. Biol. Chem. 259(21):13560-13566 (1984).
- W. Ross, T. Rowe, B. Glisson, J. Yalowich, and L. Liu. Role of topoisomerase II in mediating epipodophyllotoxin-induced DNA. Cancer Res. 44:5857-5860 (1984).
- T. Rowe, G. Kuppfer, and W. Ross. Inhibition of epipodophyllotoxin cytotoxicity by interference with topoisomerasemediated DNA cleavage. *Biochem. Pharmacol.* 34(14):2483– 2494 (1985).
- Y. S. Klausner and M. Bodansky. Coupling regents in peptide synthesis. Synthesis 453-463 (1972).
- K. H. Lee, Y. Imakura, M. Haruna, S. A. Beers, L. S. Thurston, H. J. Dia, C. H. Chen, S. Y. Liu, and Y. C. Chen. Antitumor agents. 107. New cytotoxic 4-alkylamino analogues of 4'-demethylepipodophyllotoxin as inhibitors of human DNA topoisomerase II. J. Nat. Prod. 52:606-613 (1989).
- J. H. Brewster and C. J. Ciotti, Jr. Dehydrations with aromatic sulfonyl halides in pyridine. A convenient method for the preparation of esters. J. Am. Chem. Soc. 71:6214–6215 (1955).